Preparation and characterization of nanocomposite from fresh green *Asparagus Officinalis* L. Stems and study of its biological efficacy in treating polycystic ovary syndrome induced by metformin suppository

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In this research, a green nanocomposite was meticulously prepared and characterized. Its biological effectiveness was thoroughly evaluated before and after loading it with metformin in female white rats induced with polycystic ovary syndrome (PCOS). The green nanocomposite, derived from fresh, soft asparagus stalks, underwent rigorous preparation and was carefully characterized. The nanocomposite was meticulously created by mixing 20% fresh asparagus plant with 80% silver nitrate solution (AgNO₃), and then the treatment was loaded onto it and diagnosed. Rigorous analysis involving Fourier transform infrared diffraction spectroscopy (FT-IR) and Atomic Force Microscope (AFM) images contributed to unveiling significant changes in the nanocomposite surface loaded with the treatment, all within nanoscale sizes and dimensions consistent with the results of the infrared spectrum (FT-IR). The therapeutic treatments' impact on immune parameters was comprehensively studied using 25 female white rats divided into five groups. The study conclusively indicated a significant increase (P<0.05) in the levels of cytokines IL-10 and IL-17 in the PCOS induction group (G2) compared to the control group (G1). Additionally, a significant decrease (P<0.05) in the levels of cytokine IL-10 in all treatment groups (G3-Meto., Asp.NPs-G4, G5- Asp.NPs+Meto.) compared to the PCOS induction group (G2), and a decrease (P<0.05) in the level of IL-17 in all treatment groups (G3-Meto., Asp.NPs-G4, G5- Asp.NPs+Meto.) compared with the PCOS induction group (G2) were noted. The evidence was compelling and led to the conclusion that the nano-asparagine complex was successfully prepared and displayed superior effectiveness in treating polycystic ovary syndrome compared to metformin. Furthermore, when metformin is loaded onto the nanoasparagine complex, its therapeutic effectiveness increases, reducing treatment time, side effects, and financial costs associated with repeated treatment

Keywords: Preparation of green nanocomposites; Diagnostic tests for nanocomposites; Metformin drug; Fresh green *Asparagus Officinalis* L. stems; Polycystic ovary syndrome

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INTRODUCTION

The process of synthesizing nanoparticles using plants offers several advantages over other biological methods. It eliminates the need to maintain cell density, prevents contamination from other microbial growth, and is a simple one-step synthesis method without the risk of mutations found in microorganisms. Additionally, the extraction and separation process can be easily scaled up for large-scale synthesis of nanoparticles [1,2]. Compared to microorganisms, producing plant extracts tends to be less expensive [3]. The green synthesis of nanoparticles using plant compounds as bioreducers is gaining momentum, with various plant materials such as leaf, fruit, bark, fruit peel, root, and callus extracts being utilized [4,5]. The extract contains flavonoids, terpenoids, and alkaloids that are said to help reduce and stabilize nanoparticles [6,7]. The use of nanoparticles in drug delivery technology offers several advantages. These include the ability to easily control the size and surface properties of nanoparticles to effectively deliver drugs to specific targets in the body, thereby increasing therapeutic efficiency while reducing side effects. Additionally, nanoparticles allow for controlled drug release and enable more accurate and rapid targeting of specific sites [8,9].

Polycystic Ovary Syndrome (PCOS) Studies have shown that about 8-13% of women of reproductive age and about 3-11% of adolescent girls suffer from PCOS. This syndrome is a chronic, complex, multisystem disorder and is one of the most well-known endocrine disorders that causes constant anxiety for affected women [10,11]. PCOS is a diverse disorder with different health effects and clinical symptoms that can appear in childhood and change as a person enters adolescence and adulthood [12]. There are numerous significant consequences of PCOS, including obesity, metabolic syndrome, impaired glucose tolerance, type 2 diabetes, hirsutism, acne, cardiovascular risk factors, anovulation, fertility issues, pregnancy complications, depression, anxiety, eating disorders, and a reduced quality of life [13-19]. PCOS also imposes a substantial financial burden. In the United States, the annual cost of treating the many associated health issues, such as hirsutism, infertility, diabetes, and obesity, is estimated to be approximately \$4 billion [20,21].

Metformin, also known as 1,1-dimethylbiguanide hydrochloride, is an oral hypoglycemic drug used to treat

type 2 diabetes [22,23], polycystic ovary syndrome (PCOS) [24], and other conditions such as insulin resistance in the liver. It works by inhibiting hepatic glucose production and increasing glucose uptake and utilization in peripheral tissues, such as muscle cells [25-28]. Metformin has the potential to reduce body weight [22,29] and induce ovulation [22,30]. Moreover, studies have suggested that metformin inhibits steroidogenesis in the ovary and reduces androgen production from the ovarian theca cells [22,31,32]. This makes it an important drug for the treatment of PCOS, even though the treatment process may take a considerable amount of time. There is also a good physiological basis to believe that by suppressing insulin levels through the use of insulin-sensitizing agents like metformin, it can alleviate the negative effects of ovarian stimulation and improve treatment outcomes such as ovulation and pregnancy rates [25,27,33]. Additionally, metformin may also act directly on ovarian cyst cells, reducing androgen production [25,32,34].

Cytokines are a diverse group of small proteins, typically ranging from 5-20 kDa in size [35]. They act as regulatory proteins and are secreted by cells of the immune system. These soluble, low molecular weight proteins play a crucial role in modulating the balance of humoral and cellular immunity. Additionally, they are involved in a wide range of biological activities, particularly in regulating and activating the growth, differentiation, and activation of immune cells [36,37]. Cytokines are produced by various immune cells, including B-lymphocytes, activated T-lymphocytes, macrophages, mast cells, natural killer (NK) cells, fibroblasts, and endothelial cells. It's important to note that cytokines are distinct from hormones or growth factors and function differently compared to hormones. These proteins are usually produced by specific cells [36]. Cytokines exert their effects by binding to specific receptors on target cells. The expression of these receptors on the target cells determines the amount and type of response induced by the cytokines [38]. The immune system consists of various pathways that involve proinflammatory cytokines such as TNF- α , IFN- γ , IL-1, IL-2, IL-6, IL-8, IL-12, and IL-17, as well as anti-inflammatory cytokines like IL-4, IL-10, and TGF-B. These cytokines play a crucial role in the development of autoimmune diseases. The balance between pro-inflammatory and antiinflammatory cytokines in the immune system determines how T-cells respond during an immune reaction. Proinflammatory cytokines contribute to the onset and spread of autoimmune inflammation, whereas anti-inflammatory cytokines aid in reducing inflammation and promoting recovery from the acute phase of the disease. This process is influenced by the interaction between T-helper 1 and T-helper 2, and has been a major focus in studies on cytokines and autoimmunity [39,40].

MATERIALS AND METHODS

Synthesis of silver nanoparticles using aqueous extract of fresh asparagus stems

Silver nanoparticles were synthesized by dissolving 1

millimolar of silver nitrate in 100 ml of non-ionic water at a temperature of 80 °C. The aqueous extract was prepared by dissolving 2.5 g of dried asparagus extract in 100 ml of non-ionic distilled water. Next, 20 ml of the asparagus extract was added to 80 ml of the dissolved silver nitrate solution, with a volume ratio of 80:20 (silver nitrate solution: plant extract) and at a pH of 7. The color change was observed, and the glass tubes were covered with thin silicone paper and kept in the dark in a refrigerator for five days until the color stabilized. The stabilized solution was then used in the experiment and dosed to the experimental animals once a day [41].

Glucophage drug (Metformin)

Metformin is produced by Merck Sante s.a.s2, rue du Pressoir Vert-45400 SEMOY-France. Each box contains 5 strips, with each strip containing 20 tablets and each tablet containing 500 mg. This 500 mg is one of the therapeutic doses used for humans. The required therapeutic dose was prepared in the study based on the equation for converting the therapeutic dose from humans to animals, as calculated according to [42].

Loading the drug metformin on the green nano-asparagus compound

To load the drug metformin onto the green nanoasparagus compound, start by dissolving 1250 mg of metformin in 40 ml of deionized water on a hot plate magnetic stirrer. Then, take a graduated glass beaker and add 40 ml of the green nano-complex to it. Next, add the dissolved metformin to the beaker. Cover the beaker with silicone paper and leave it on the hot plate magnetic stirrer (not exceeding 50 degrees Celsius) until the mixture reaches a volume of 13.33. After reaching this volume, add more of the green nano-complex until the volume reaches 40 ml again. Return the mixture to the device for 10 minutes to ensure homogeneity, and then store it in the refrigerator until it is used in the experiment [43,44].

Study groups

The study involved 25 female white rats aged 5-7 months, weighing between 165-175 grams. The rats were randomly divided into four groups for two treatment periods, with each group comprising five females. The groups were treated as follows: Negative control group (G1): Five female white rats were dosed with physiological saline (0.9%), i.e., normal saline only. Positive control/induction group (G2): Five female white rats were orally dosed with 1mg of letrozole dissolved in 1% Carboxymethylcellulose powder (CMC) dissolved in 99% distilled water for 28 days to induce polycystic ovary syndrome. Group treated with metformin only (G3): Five female albino rats with induced polycystic ovary syndrome were treated with oral metformin at a concentration of 500 mg/kg body weight for 28 days. Group treated with nano-asparagine only (G4): Five female rats with induced polycystic ovary syndrome were treated with oral doses of nano-asparagine at a concentration of 300 mg/kg of body weight for 28

days. Group treated with nano-asparagine loaded with metformin (G7): Five female albino rats with induced polycystic ovary syndrome were treated with oral doses of nano-asparagine loaded with metformin at a concentration of 300 mg/kg of body weight for 28 days.

Collection of blood samples

Blood samples were taken from the animals that were both treated and untreated after being induced with the disease. Each animal's blood, drawn directly from the heart by heart puncture after being anesthetized with chloroform, was collected at a rate of 3 ml using sterile medical syringes and placed in gel tubes without any anticoagulant. The tubes were left for 15-20 minutes at room temperature for the blood to clot before being centrifuged at 3000 rpm for 5-10 minutes to separate the blood serum. The sera were transferred to 1.2 ml Eppendorf tubes and stored in the refrigerator at 20 °C to conduct immune tests, which included assessing inflammatory and anti-inflammatory interleukins (IL-10 and IL-17) [45].

Immune tests estimation

Method of measuring IL-17 and IL-10 concentration

- 1. All reagents were prepared and placed on a clean place at room temperature.
- 2. The plate was fixed on a flat place.
- 3. 100 microliters of incubation buffer were added using a micropipette to each well.
- $\ 4. \ \ 100 \ \ \mu l \ \ of \ \ Calibrators \ (Control) \ \ solution \ \ Samples \ \ were \ added \ to \ \ each \ well.$
- 5. 50 µl of conjugate solution was added to each well.
- 6. Incubated for 2 h at room temperature on a shaker at 700 rpm = 100 rpm.
- 7. Excess liquid was removed from each well.
- 8. The plate was washed three times by:
 - Adding 0.4 ml of wash solution to each well.
 - Removing all components from the wells.
- 9. Adding 200 μ l of freshly prepared substrate solution to each well for 15 min followed by washing the wells.
- 10. The plate was incubated for 30 min at room temperature on a shaker at 700 rpm +100 rpm in the dark.
- 11.50 μl of Stop solution was added to each well.

12. The absorbance spectrum was read at a wavelength of 450 nm.

RESULTS AND DISCUSSION

Identification of metformin, asparagus stem extract and asparagus nanocomposite before and after loading with metformin.

Infrared Spectroscopy (FT-IR)

FT-IR is a crucial tool for identifying active functional groups. The FT-IR spectra of AgNPs synthesized using an aqueous extract of fresh green asparagus stalks are provided, indicating the groups responsible for covering and stabilizing the nanoparticles [46]. These techniques are used to study the secondary structure and assembly of proteins, as well as the composition of carbohydrates and nucleic acids. The resulting curves show the chemical groups and help identify the active compounds in the extract that contribute to the oxidation and reduction processes involved in producing nanoparticles [47]. To identify the active functional groups in the aqueous extract of fresh green asparagus stalks responsible for reducing silver nitrate to silver nanoparticles and stabilizing them, an infrared spectrometer equipped with Fourier transform (FTIR) should be used. The FTIR analysis results, shown in Fig. 1. 2. and 3., revealed different bond extensions at various wave numbers. The AgNPs synthesized using the aqueous extract of fresh green asparagus stalks exhibited overlapping bands ranging between 1076.32-1500 cm⁻ ¹. In contrast, the metformin drug displayed bands at frequencies of 934.91-1497.82 cm⁻¹. When examining the nano asparagus complex loaded with metformin drug, we observed a shift in frequencies to 2074.85, 3437.20, and 1637.71 cm⁻¹, indicating the presence of stretching of the hydroxyl groups O-H bonds found in alcohols and phenols.

The AgNPs composite silver nanoparticles showed a broad absorption peak at a frequency of 3437.02 cm⁻¹, which is attributed to the N-H amine group stretching vibration. In comparison, the free metformin drug exhibited an absorption peak at a frequency of 3436.87 cm⁻¹, also indicating the N-H amine bond stretching vibration. The infrared spectrum of the drug-loaded nanocomposite displayed a shift towards a frequency of 3437.20 cm⁻¹, suggesting the N-H amine bond stretching. Furthermore, in the nano-asparagine complex, the bands appeared at frequencies between 1000 - 1500 cm⁻¹, indicating the vibration of the C-O bond, which exhibits a strong extension. In the analysis of the metformin drug, we observed frequency bands between 1050.38-1497.82 cm⁻¹, which are attributed to the C-N bond and exhibit strong extension. When examining the nano-composite loaded with the drug, we noticed a shift of the band at 1637.71 cm⁻¹compared to the examination of the nano-asparagine complex and the metformin drug. However, the band at



2074.85 cm⁻¹ remained at almost the same frequencies. The less broad peak of 1637.54 cm⁻¹ in the asparagine nanocomposite test indicates the extension of the C=C bond in the alkene group. This differs from the free drug test, in which the peak of 1638.30 cm⁻¹ resulted from the C=C bond. In the test of the asparagine nanocomposite loaded with the drug, we observed that the peak remained at the same frequency of 1637.71 cm⁻¹. The infrared spectrum of the nanocomposite loaded with the drug showed a shift and stretching of the C-O bond in the frequency range of 1000-1500 cm⁻¹, following the results of the test for the asparagine nanocomposite and the metformin drug test, which were at the frequencies of 1050.38 and 1076.32 cm⁻¹, respectively. The low peaks observed in the nano-asparagus, drug, and drug-loaded nanocomposite at frequencies of

690.90, 696.02, and 688.38 cm⁻¹, respectively, within the C-H bond, showed strong stretching of alkanes. There was a noticeable shift in the results between the drug and drug-loaded nanocomposite examinations. The vibrational peaks (absorption and transmittance) were consistent with the presence of flavonoids, proteins, and terpenoids in the aqueous extract of fresh green asparagus stems. These compounds are likely responsible for the synthesis and reduction of nanoparticles, as reported by Posinastty, et al. [48] and Rojas and Asmat-Campos [49]. The presence of organic functional groups, such as alkanes and alcohols, played an important role in reducing silver nitrate and forming silver nanoparticles. These groups also helped prevent the particles from oxidation and deterioration, contributing to their long-term stability [50,51]. The

displacements observed in the drug-loaded nanocomposite provide clear evidence of metformin being loaded onto the asparagine nanocomposite.

Atomic Force Microscopy (AFM)

In Fig. 4. and 5., the current study presented microscopic images captured by the atomic force microscope. The roughness coefficient of the outer surface of the asparagus nanocomposite particles was 0.99681 nm, while the surface roughness coefficient of the aqueous extract of fresh green asparagus stalks was 1.42235 nm. The difference before and after converting the aqueous extract of fresh green asparagus stalks to the nanocomposite was 0.42554 nm, signaling the success of the process of preparing the nanocomposite from the aqueous extract of green asparagus. In comparison, the roughness coefficient of the free metformin drug was 1.9006 nm. Furthermore, the square root rate of the asparagus nanocomposite was 1.28600 nm, whereas the aqueous extract of fresh green asparagus stalks measured 1.84616 nm. The difference in the square root rate was 0.56016 nm, implying that the nanocomposite falls within the nanoscale limits [43]. The square root of the free Metformin was 2.36988 nm. The surface area of the asparagus nanocomposite was 3.113%, and the surface area of the aqueous extract of fresh green asparagus stalks was 5.019%. This indicates that the surface is a good receptor for the particles to be loaded. The surface area of the free metformin was 7.441%. The maximum peak height of the asparagus nanocomposite was 9.9939, and the maximum peak height of the aqueous extract of fresh green asparagus stalks was 11.7714. The maximum peak height of the free metformin was 13.6780. The maximum pit height of the asparagus nanocomposite was 4.5740, and the maximum pit height of the aqueous extract of fresh green asparagus stalks was 11.4198. The maximum pit height of the free metformin was 10.9336. The surface luminous intensity values of the asparagus nanocomposite were 2.186, and the surface luminous intensity values of the fresh green asparagus extract were 2.857. The surface light intensity values of free metformin were 3.531. The study's results revealed that the average diameter of the Asp.NPs nano asparagus compound particles was 45.83 nm. This was in contrast to the average diameter of the green asparagus extract Asp. particles, which measured 116.9 nm. The difference between the two average sizes was 71.07 nm. Additionally, the average diameter of the free metformin particles was 176.5 nm, as indicated in Tab. 1. Fig. 6. of the study showed that the nano-asparagine complex loaded with metformin had a roughness coefficient of 3.03736 nm, while the roughness coefficient of free metformin was 1.90062 nm. The difference in the roughness coefficient was 1.13674 nm. This is a significant finding as it ensures that the nanoasparagine complex is properly loaded with metformin following the conversion of fresh green asparagus into the



Tab. 1. Physical properties of metformin, asparagus stem extract and green asparagus nanocomposite before and after loading with metformin.

Physical properties of therapeutic agents	Meto. (nm)	Asper. (nm)	Asper.Nps (nm)	AsperNps / Meto. (nm)
Maximum pit height (Sv)	10.9336	11.4198	4.7540	23.6796
Kurtosis (Sku)	3.531	2.857	2.186	3.793
Developed interfacial area ratio (Sdr)	7.441%	5.019%	3.113%	27.80%
Maximum peak height (Sp)	13.6780	11.7714	9.9939	27.6571
Arithmetical mean height (Sa)	1.90062	1.42235	0.99681	3.03736
Root mean square height(Sq)	2.36988	1.84616	1.28600	4.34698
Mean size	116.9	176.5	45.83	278.4

Fig. 6. Shows a three-dimensional image of the atomic force microscope of the Asparagine Nanocomposite loaded with Metformin (Asp.NPs./ Meto.).



nano-complex [52]. The square root mean of the nanoasparagine complex loaded with metformin was 4.34698 nm, while that of metformin alone was 2.36988 nm, with the difference in the square root mean being 1.9771 nm. Additionally, the surface area of the nano-asparagine complex loaded with metformin was 27.80%, whereas the surface area of free metformin was 7.441%. The results also indicated that the surface luminescence intensity values of the metformin-loaded nano-asparagine were 3.793, while those of the free metformin were 3.531. Furthermore, the maximum peak height of the metformin-loaded nanoasparagine was 27.6571, in contrast to the 13.6780 value of the free metformin. The maximum pit height of the metformin-loaded nano-asparagine was 23.6796, while that of the free metformin was 10.9336. The average particle size of the metformin-loaded nano-asparagine Asp.NPs/Meto. Was 278.4 nm. This is larger than the average particle size of metformin alone, which was 176.5 nm, and the nano-asparagine, which was 45.83 nm. This demonstrates the successful loading of metformin. This information is summarized in Tab. 1., which illustrates the physical properties of metformin particles, fresh green asparagus stem extract, and the asparagus nanocomposite before and after loading with metformin.

Study the effect of induction of polycystic ovary syndrome and therapeutic treatments on some immune parameters (IL-10) and (IL-17) in the blood serum of female rats

The results of the current study, as shown in **Tab. 2.**, revealed different levels of immune parameters in the various groups. It is important to note that there was a

group with induced PCOS (G2) compared to the control group (G1). Additionally, there was a significant increase (P<0.05) in the level of IL-10 in the group treated with Meto (G3) compared to the control group (G1). Furthermore, the results indicated a significant increase (P<0.05) in the level of IL-10 in the group treated with Asp.NPs. (G4) compared to the control group (G1). In contrast, the group treated with Asp.NPs./Meto (G5) showed an increase in the level of IL-10 that did not reach statistical significance compared to the control group (G1). The results from Tab. 1. also demonstrated a decrease in the level of IL-10 in the group treated with Meto. (G3) compared to the PCOS induction group (G2), although it did not reach statistical significance. Moreover, there was a significant decrease (P<0.05) in the level of IL-10 in the group treated with Asp.NPs. (G4) compared to the PCOS induction group (G2). Similarly, there was a significant decrease (P<0.05) in the level of IL-10 in the group treated with Asp.NPs./ Meto (G5) compared to the PCOS induction group (G2).

significant increase (P<0.05) in the level of IL-10 in the

The results from **Tab. 2.** show varying levels of immune parameters in the studied groups. There is a significant increase (P<0.05) in the level of IL-17 in the PCOS induction group (G2) compared to the control group (G1). Additionally, there is a significant increase (P<0.05) in the level of IL-17 in the group treated with Meto (G3) compared to the control group (G1). The results also demonstrate a significant increase (P<0.05) in the level of IL-17 in the group treated with Asp.NPs (G4) compared to the control group (G1), as well as in the group treated with Asp.NPs./Meto (G5) compared to the control group (G1). Furthermore, the results show a significant decrease (P<0.05) in the level of IL-17 in the group treated with Meto (G3) compared to the PCOS induction group (G2), Shaheen AA, et al. – Preparation and characterization of nanocomposite from fresh green Asparagus Officinalis L. Stems and study of its biological efficacy in treating polycystic ovary syndrome induced by metformin suppository...

Tab. 2. Shows the effect of polycystic ovary syndrome and treatment with metformin and asparagine nanocomposite before and after loading it with metformin on the levels of immune parameters (IL-10) and (IL-17) in the serum of female albino rats.	Transactions	IL-10	IL-17
	Control	0.613 ± 0.095	0.146 ± 0.740
	PCOS	1.320 ± 0.146	0.101 ± 1.213
	Meto.	1.053 ± 0.309	0.131 ± 1.048
	Asp.NPs	0.820 ± 0.096	0.097 ± 0.941
	Asp.NPs/ Meto.	0.694 ± 0.115	0.247 ± 0.994
	L.S.D.	0.275	0.202

as well as in the group treated with Asp.NPs (G4) and Asp. NPs./Meto (G5) compared to the PCOS induction group (G2).

The results of the current study revealed that the levels of interleukin-10 (IL-10) were elevated above normal in the PCOS group. This increase is attributed to heightened inflammation in the body, which stimulates the main anti-inflammatory white blood cells. IL-10 functions to suppress the activities of monocytes, macrophages, and pro-inflammatory cytokines like IL-17, IL-10, TNF-a, neutrophils, and T cells. Furthermore, interleukin-10 is linked to obesity and works to counteract pro-inflammatory cytokines that contribute to insulin resistance in women with polycystic ovary syndrome. IL-10 serves as the primary anti-inflammatory cytokine regulating the immune system. According to Al-Hashem [53], treatment with Meto led to a decrease in the level of interleukin-10. This is attributed to the drug's effectiveness in regulating sugar, insulin, and cholesterol levels, which in turn reduces obesityrelated inflammation. Additionally, the drug is effective against diabetes and inflammation. As a result, it works to reduce the level of interleukin-10, which acts as an antiinflammatory and helps alleviate symptoms of polycystic ovary syndrome [54]. Furthermore, the anti-inflammatory effects of fresh asparagus stem extract used in the green nanocomposite preparation can reduce the production of IL-17, IL-6, and TNF-a, while causing an increase in IL-10 in the blood [55]. The therapeutic potential of nanocomposites has led to their widespread use in recent years. When the aqueous extract of fresh asparagus stems is used to prepare green nanocomposites, it increases the effectiveness of asparagine acid present in the extract against inflammation. This results in the production of effective doses of IL-10, which prevents the synthesis of pro-inflammatory cytokines [56]. In various studies, the effect of metformin on body mass index and insulin sensitivity has been observed. It is suggested that there may be a synergistic relationship between the chemical compounds in metformin, which can reduce insulin and lipid levels in the body, enhance glucose absorption, and consequently reduce body mass [57]. We observed a significant increase in the effectiveness of metformin when it was loaded onto the green nanocomposite. This led to positive therapeutic outcomes by reducing the levels of anti-inflammatory interleukins compared to the stimulated group. The enhanced effectiveness is attributed to the green nanosilver compounds, which target inflammatory areas in the body. This combined treatment approach not only reduces obesity-related inflammation but also lowers oxidative stress, thereby reducing anti-inflammatory agents in the body [55,58].

The findings of the recent study revealed that the level of interleukin-17 (IL-17) was elevated above normal in the PCOS group. This increase is attributed to heightened levels of oxidative stress, insulin resistance, ovarian dysfunction, inflammation, obesity, and other characteristics associated with polycystic ovary syndrome. The study also observed the presence of subsets of inflammatory T helper 17 (Th17) cells that produce both IL-17A and IL-17F, and these cells depend on signals generated by high levels of both IL-6 and TNF-a [59]. Stimulation of TGF-B cells with IL-6 leads to increased expression of IL-21 and activation of signal transducer and activator of transcription-3. This activation then leads to the activation of Th17 subsets and drives the proinflammatory IL-17 response [60,61]. Similarly, TNF-a may also cooperate with IL-23 to promote IL-17A expression [62]. Additionally, TNF-a indirectly promotes IL-17A by inducing IL-6 expression [59,63]. Hence, higher levels of IL-6 and TNF- α in females with PCOS may influence the expression of IL-17 cytokines, which could contribute to the development of the disease. Additionally, proinflammatory members of the IL-17 family might also add to the complexity of the inflammatory response in PCOS by working together with TNF- α to stimulate IL-6 expression. IL-17 cytokines have been found to play a significant role in PCOS-related complications such as joint stiffness, a major contributor to cardiovascular disease [64]. Additionally, they have synergistic effects with TNF- α on oxidative stress [59]. Metformin reduces insulin resistance, lipid profile, and cholesterol levels, while enhancing glucose absorption. This helps regulate these factors and reduce inflammation in the body, which has increased due to obesity and oxidative stress [65]. Asparagine nanocomposite Asp.NPs have shown anti-inflammatory activity by reducing the release of pro-inflammatory cytokines (TNF-alpha, IL-6, IL-1 beta, and IL-17). Anti-inflammatory cytokines are regulatory molecules that inhibit the immune response [66]. Nanoparticles can be designed to directly target immune cells and suppress their activity or avoid immune recognition [67]. Aspartate nanoparticles (Asp.NPs) are important in inhibiting inflammatory promoters such as cytokines and proinflammatory enzymes. Many metal and metal oxide nanocomposites, such as silver, have been reported to possess anti-inflammatory properties when compared to their bulk counterparts. Utilizing green silver nanocomposites as an anti-cytokine approach is an interesting strategy to reduce inflammation, as it blocks the interaction between cytokines and their receptors. Polycystic ovary syndrome (PCOS) is a pro-inflammatory condition, and Asp.NPs has a beneficial effect in reducing

inflammatory cytokines [68]. The main advantage of nano-drug carriers, such as the metformin-loaded asparagus stem nanocomposite, is to improve the targeted delivery of drugs to the intended site of action. This delivery process is referred to as drug targeting and can be achieved through active or passive targeting methods. By encapsulating biologically active molecules in nanocarriers, their bioavailability can be increased, leading to enhanced stability. Utilizing nanocarriers in drug delivery enhances the quality of drugs, thereby reducing side effects and the required drug dosage. Consequently, this drug, when administered after suppository, has a greater impact on reducing inflammation-causing elements (IL-17 and IL-6) [69,70].

CONCLUSION

This study demonstrates that the green nanocomposite was successfully prepared from fresh asparagus stems based on the results obtained from AFM and FTIR examination. The study also showed that the loading of metformin on the asparagus nanocomposite was successful, as confirmed by AFM and FTIR examination results. Additionally, it was concluded that the effectiveness of the asparagus nanocomposite, both before and after loading with metformin, in treating polycystic ovary syndrome was high compared to the free metformin drug, and it achieved results in a shorter period of time. Consequently, this bioprepared nanocomposite reduced side effects, treatment cost, and recovery time.

- Amargo MM, Bucoya EA, Fundador EO, et al. Plant-mediated synthesis of silver nanoparticles using mangosteen pericarp extract and their antimicrobial potential. *Nanoscience & Nanotechnology-Asia*. 2023;13(2):64-71.
 Veerasamy R, Xin TZ, Gunasagaran S, et al. Biosynthesis of silver
 - Veerasamy R, Xin TZ, Gunasagaran S, et al. Biosynthesis of silver nanoparticles using mangosteen leaf extract and evaluation of their antimicrobial activities. J Saudi Chem Soc. 2011;15(2):113-120.
 - Mittal AK, Chisti Y, Banerjee UC. Synthesis of metallic nanoparticles using plant extracts. *Biotechnol Adv.* 2013;31(2):346-356.
 - Pandit C, Roy A, Ghotekar S, et al. Biological agents for synthesis of nanoparticles and their applications. J King Saud Univ Sci. 2022;34(3):101869.
 - Rai M, Yadav A. Plants as potential synthesiser of precious metal nanoparticles: Progress and prospects. *IET Nanobiotechnol.* 2013;7(3):117-124.
 - Halimi M, Nasrabadi M, Soleamani N, et al. Green synthesis of nanosilver particles from extract of Dracocephalum Lindbergii. *Asian J Nano Mat.* 2018;1(1):19-24.
 - Dubey M, Bhadauria S, Kushwah BS. Green synthesis of nanosilver particles from extract of *Eucalyptus* hybrida (safeda) leaf. *Dig J Nanomater Biostruct*. 2009;4(3):537-543.
 - Mohanraj M, Ayyannan G, Raja G, et al. Synthesis, spectral characterization, DNA interaction, radical scavenging and cytotoxicity studies of ruthenium (II) hydrazone complexes. J Photochem. Photobiol B. 2016;158:164-173.
 - Mohanraj VJ, Chen YJ. Nanoparticles-a review. Trop J Pharm Res. 2006;5(1):561-573.
 - Naz MS, Tehrani FR, Majd HA, et al. The prevalence of polycystic ovary syndrome in adolescents: A systematic review and metaanalysis. Int J Reprod Biomed. 2019;17(8):533.
 - Bozdag G, Mumusoglu S, Zengin D, et al. The prevalence and phenotypic features of polycystic ovary syndrome: A systematic review and meta-analysis. *Hum Reprod.* 2016;31(12):2841-2455.
 - Teede H, Deeks A, Moran L. Polycystic ovary syndrome: A complex condition with psychological, reproductive and metabolic manifestations that impacts on health across the lifespan. BMC Med. 2010;8:1-0.
 - Lim SS, Davies MJ, Norman RJ, et al. Overweight, obesity and central obesity in women with polycystic ovary syndrome: A systematic review and meta-analysis. *Hum Reprod Update*. 2012;18(6):618-637.
 - Joham AE, Boyle JA, Zoungas S, et al. Hypertension in reproductiveaged women with polycystic ovary syndrome and association with obesity. *Am J Hypertens.* 2015;28(7):847-851.
 - Dokras A, Stener-Victorin E, Yildiz BO, et al. Androgen excesspolycystic ovary syndrome society: position statement on depression, anxiety, quality of life, and eating disorders in polycystic ovary syndrome. *Fertil Steril.* 2018;109(5):888-899.
 - 16. Kakoly NS, Khomami MB, Joham AE, et al. Ethnicity, obesity and the prevalence of impaired glucose tolerance and type 2 diabetes

in PCOS: A systematic review and meta-regression *Hum Reprod Update*. 2018;24(4):455-467.

- Tay CT, Teede HJ, Hill B, et al. Increased prevalence of eating disorders, low self-esteem, and psychological distress in women with polycystic ovary syndrome: A community-based cohort study. *Fertil Steril.* 2019;112(2):353-361.
- Lim SS, Kakoly NS, Tan JW, et al. Metabolic syndrome in polycystic ovary syndrome: A systematic review, meta-analysis and metaregression. Obes Rev. 2019;20(2):339-352.
- Tay CT, Teede HJ, Loxton D, et al. Psychiatric comorbidities and adverse childhood experiences in women with self-reported polycystic ovary syndrome: An Australian population-based study. *Psychoneuroendocrinology*. 2020;116:104678.
- Maqbool M, Gani I, Geer MI. Polycystic ovarian syndrome-a multifaceted disease: A review. Int J Pharm Sci Res. 2019;10(3):1072-1079.
- Azziz R, Marin C, Hoq L, et al. Health care-related economic burden of the polycystic ovary syndrome during the reproductive life span. J Clin Endocrinol Metab. 2005;90(8):4650-8.
- 22. Furat Rencber S, Kurnaz Ozbek S, Eraldemir C, et al. Effect of resveratrol and metformin on ovarian reserve and ultrastructure in PCOS: an experimental study. J Ovarian Res. 2018;11:1-6.
- 23. Kai Y, Kawano Y, Yamamoto H, et al. A possible role for AMPactivated protein kinase activated by metformin and AICAR in human granulosa cells. *Reprod Biol Endocrinol.* 2015;13:1-8.
- 24. Patel R, Shah G. Effect of metformin on clinical, metabolic and endocrine outcomes in women with polycystic ovary syndrome: A meta-analysis of randomized controlled trials. *Curr Med Res Opin.* 2017;33(9):1545-1557.
- Tso LO, Costello MF, Albuquerque LE, et al. Metformin treatment before and during IVF or ICSI in women with polycystic ovary syndrome. Cochrane Database Syst Rev. 2020(12).
- Nardo LG, Rai R. Metformin therapy in the management of polycystic ovary syndrome: Endocrine, metabolic and reproductive effects. *Gynecol Endocrinol.* 2001;15(5):373-380.
- Dunaif A, Segal KR, Futterweit W, et al. Profound peripheral insulin resistance, independent of obesity, in polycystic ovary syndrome. *Diabetes*. 1989;38(9):1165-1174.
- Barbieri RL, Makris A, Randall RW, et al. Insulin stimulates androgen accumulation in incubations of ovarian stroma obtained from women with hyperandrogenism. J Clin Endocrinol Metab. 1986;62(5):904-910.
- Ladson G, Dodson WC, Sweet SD, et al. The effects of metformin with lifestyle therapy in polycystic ovary syndrome: A randomized double-blind study. *Fertil Steril*. 2011;95(3):1059-1066.
- 30. Palomba S, Orio Jr F, Falbo A, et al. Prospective parallel randomized, double-blind, double-dummy controlled clinical trial comparing clomiphene citrate and metformin as the firstline treatment for ovulation induction in nonobese anovulatory women with polycystic ovary syndrome. J Clin Endocrinol Metab. 2005;90(7):4068-4074.

- **31.** Mansfield R, Galea R, Brincat M, et al. Metformin has direct effects on human ovarian steroidogenesis. *Fertil Steril.* 2003;79(4):956-962.
- Attia GR, Rainey WE, Carr BR. Metformin directly inhibits androgen production in human thecal cells. *Fertil Steril.* 2001;76(3):517-524.
- 33. Tang T, Barth JH, Balen AH. Effect of metformin on follicular anti-Mullerian hormone concentrations in women with PCOS undergoing IVF treatment. InHuman Reproduction, Great clarendon st, oxford ox2 6dp, england: oxford univ press 2010; Jun 1: 168-168.
- **34.** Palomba S, Falbo A, Russo T, et al. Systemic and local effects of metformin administration in patients with Polycystic Ovary Syndrome (PCOS): Relationship to the ovulatory response. *Hum Reprod.* 2010;25(4):1005-1013.
- 35. Ibelgaufts H. Cytokines and cells online pathfinder encyclopedia Version 31.4. Spring. Summer.
- Chokkalingam V, Tel J, Wimmers F, et al. Probing cellular heterogeneity in cytokine-secreting immune cells using dropletbased microfluidics. *Lab Chip.* 2013;13(24):4740-4744.
- Kleiner G, Marcuzzi A, Zanin V, et al. Cytokine levels in the serum of healthy subjects. *Mediators Inflamm*. 2013;2013(1):434010.
- Angiolilli C, Grabiec AM, Ferguson BS, et al. Inflammatory cytokines epigenetically regulate rheumatoid arthritis fibroblastlike synoviocyte activation by suppressing HDAC5 expression. Ann Rheum Dis. 2016;75(2):430-438.
- Shachar I, Karin N. The dual roles of inflammatory cytokines and chemokines in the regulation of autoimmune diseases and their clinical implications. J Leukoc Biol. 2013;93(1):51-61.
- 40. Romero-Adrián TB, Leal-Montiel J, Monsalve-Castillo F, et al. Helicobacter pylori: Bacterial factors and the role of cytokines in the immune response. Curr Microbiol. 2010;60:143-155.
- Nahar K, Aziz S, Bashar M, et al. Synthesis and characterization of Silver nanoparticles from Cinnamomum tamala leaf extract and its antibacterial potential. Int J Nano Dimens. 2020;11(1):88-98.
- Al- Safi, A. H. M. Effect of Dexamethasone on blastocyst implantation, Cartilage ,bone embryo and some serum biochemical parameters in pregnant rats . 2018; 150.
- 43. Marza Hamza N, M Hussain KA, Al-Safy AH. Synthesis of nanoscale xerogel/MTX and study its effects on the liver and kidney tissue and level of IgG in rats with rheumatoid arthritis. J Nanostruct. 2022;12(2):254-261.
- Wang Y, Li C, Wan Y, et al. Quercetin loaded ceria nanocomposite potentiate dual directional immunoregulation via macrophage polarization against periodontal inflammation. Small. 2021;17(41):2101505.
- Pinon-Lataillade G, Thoreux-Manlay A, Coffigny H, et al. Reproductive toxicity of chronic lead exposure in male and female mice. *Hum Exp Toxicol.* 1995;14(11):872-878.
- **46. Huq MA.** Green synthesis of silver nanoparticles using Pseudoduganella eburnea MAHUQ-39 and their antimicrobial mechanisms investigation against drug resistant human pathogens. *Int J Mol Sci.* 2020;21(4):1510.
- Ami D, Mereghetti P, Natalello A. Contribution of infrared spectroscopy to the understanding of amyloid protein aggregation in complex systems. *Front Mol Biosci.* 2022;9:822852.
- Posinasetty B, Kommineni S, Kumarachari RK, et al. Design And optimization of nano encapsulated bio compounds of *Asparagus racemosus*: box behnken approach. *Int J App Pharm.* 2024;16(1):134-149.
- 49. Rojas ML, Asmat-Campos D. Optimization of ultrasound-assisted extraction of bioactive compounds from asparagus (Asparagus officinalis) by-products and its application in silica nanoparticle synthesis. LACCEI. 2023;1(8).
- Ahmed T, Shahid M, Noman M, et al. Silver nanoparticles synthesized by using Bacillus cereus SZT1 ameliorated the damage of bacterial leaf blight pathogen in rice. *Pathogens*. 2020;9(3):160.
- Majeed S, Aripin FH, Shoeb NS, et al. Bioengineered silver nanoparticles capped with bovine serum albumin and its anticancer

and apoptotic activity against breast, bone and intestinal colon cancer cell lines. *Mater Sci Eng*: C. 2019;102:254-263.

- 52. Hussein KA, Khalaf AA. Preparation, diagnosis and study of the inhibitory effect of copper nanoparticles before and after Erythromycin loading on Pseudomonas aeruginosa. *IOP Conf Ser Mater Sci Eng.* 2020;928: 6.
- Al-Hashem F. Suppression of L-arginine-induced acute necrotizing pancreatitis in rats by metformin associated with the inhibition of myeloperoxidase and activation of interleukin-10. *Int J Morphol.* 2021;39(1):102-108.
- Lord JM, Flight IH, Norman RJ. Metformin in polycystic ovary syndrome: Systematic review and meta-analysis. *Bmj*. 2003;327(7421):951.
- 55. Mousavi Maleki MS, Ebrahimi kiasari R, Seyed Mousavi SJ, et al. Bromelain-loaded nanocomposites decrease inflammatory and cytotoxicity effects of gliadin on Caco-2 cells and peripheral blood mononuclear cells of celiac patients. *Sci Rep.* 2023;13(1):21180.
- Wang X, Coradin T, Hélary C. Modulating inflammation in a cutaneous chronic wound model by IL-10 released from collagen-silica nanocomposites via gene delivery. *Biomater Sci.* 2018;6(2):398-406.
- Naderpoor N, Shorakae S, de Courten B, et al. Metformin and lifestyle modification in polycystic ovary syndrome: Systematic review and meta-analysis. *Hum Reprod Update*. 2015;21(5):560-574.
- El-Demerdash FM, Al Mhanna AB, El-Sayed RA, et al. Hepatoprotective impact of Nigella sativa silver nanocomposite against genotoxicity, oxidative stress, and inflammation induced by thioacetamide. *Tissue Cell*. 2024;87:102332.
- 59. Özçaka Ö, Buduneli N, Ceyhan BO, et al. Is interleukin[17 involved in the interaction between polycystic ovary syndrome and gingival inflammation?. J Periodontol. 2013;84(12):1827-1837.
- 60. Kwok SK, Cho ML, Her YM, et al. TLR2 ligation induces the production of IL-23/IL-17 via IL-6, STAT3 and NF-kB pathway in patients with primary Sjogren's syndrome. *Arthritis Res Ther.* 2012;14:1-3.
- Bettelli E, Korn T, Kuchroo VK. Th17: The third member of the effector T cell trilogy. Curr Opin Immunol. 2007;19(6):652-657.
- 62. Liu Z, Yadav PK, Xu X, et al. The increased expression of IL-23 in inflammatory bowel disease promotes intraepithelial and lamina propria lymphocyte inflammatory responses and cytotoxicity. J Leukoc Biol. 2011;89(4):597-606.
- Brenner DA, O'Hara M, Angel P, et al. Prolonged activation of jun and collagenase genes by tumour necrosis factor-[]. Nature. 1989;337(6208):661-663.
- Butcher M, Galkina E. Current views on the functions of interleukin-17A-producing cells in atherosclerosis. *Thromb Haemost.* 2011;106(11):787-795.
- Mohammed FM, Pambuk CI, Al-Kadh NA. Evaluation of interleukin 17 A level and lipid profile in diabetic female patients treated by Metformin and Glimepiride in Kirkuk City Iraq. *Biomed Pharmacol J.* 2019;12(4):1849-1855.
- 66. Opal SM, DePalo VA. Anti-inflammatory cytokines. Chest. 2000;117(4):1162-1172.
- Kawai T, Akira S. Toll-like receptors and their crosstalk with other innate receptors in infection and immunity. *Immunity*. 2011;34(5):637-650.
- Mugade M, Patole M, Pokharkar V. Bioengineered mannan sulphate capped silver nanoparticles for accelerated and targeted wound healing: Physicochemical and biological investigations. *Biomed Pharmacother*. 2017;91:95-110.
- Yacoub AS, Ammar HO, Ibrahim M, et al. Artificial intelligenceassisted development of in situ forming nanoparticles for arthritis therapy via intra-articular delivery. Drug Deliv. 2022;29(1):1423-1436.
- Ulbrich W, Lamprecht A. Targeted drug-delivery approaches by nanoparticulate carriers in the therapy of inflammatory diseases. J R Soc Interface. 2010;7(suppl_1):S55-66.